Rhizobium Symbiosis Contribution to Short-term Salt Stress Tolerance in Alfalfa (Medicago sativa L.)

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Introduction	Results	Leaf Root Leaf A A A A A A A A A A A A A A A A A A A
Symbiotic nitrogen fixation (SNF) has profound	Shoot Root	$\begin{bmatrix} a \\ b \\ b \\ b \\ c \\ c \\ c \\ c \\ c \\ c \\ c$

agronomic, economic, and ecological impacts because availability of soil nitrogen most frequently limits agricultural production throughout the world. In SNF, plants provide carbon compounds and energy sources needed by rhizobia in exchange for fixed nitrogen. In addition to nitrogen fixation, rhizobia have been shown to benefit plants in many different ways, which imply that symbiosis may enhance the overall health status of plants and improve their tolerance to abiotic stresses, including salinity.

Aim

The objective of this study is to test our hypothesis that rhizobium symbiosis may improve alfalfa salt tolerance physiological and biochemical affecting its by processes such as enhancing antioxidant capacity during stress response. We studied the response of well-developed alfalfa plants with active nodules, inactive nodules and no nodules to salt stress by assessing survival rate and their ability to deal with oxidative and osmotic stress induced by salt shock.



Table 1 Variance analysis of survival rate, fresh weight of regenerated shoots, activities of SOD, POD, CAT and APX in the leaf and root of alfalfa under different treatments of nodulation and salt shock

_	Survival		Fresh weight		SOD		POD		CAT		APX	
DF	F	Р	F	Р	F	Р	F	Р	F	Р	F	Р
	value	value	value	value	value	value	value	value	value	value	value	value
2	5.33	0.022	8.05	0.001	2.19	0.122	39.79	< 0.001	49.79	< 0.001	37.96	< 0.001
5	133.67	< 0.001	105.78	< 0.001	9.08	< 0.001	47.51	< 0.001	23.39	< 0.001	24.16	< 0.001
10	5.33	0.007	4.08	0.001	7.50	< 0.001	21.97	< 0.001	11.84	< 0.001	7.84	< 0.001
2					15.64	< 0.001	53.41	< 0.001	56.22	< 0.001	3.57	0.035
5					15.1	< 0.001	87.34	< 0.001	5.48	< 0.001	3.08	0.016
10					6.12	< 0.001	21.22	< 0.001	2.94	0.005	4.33	< 0.001
	DF 2 5 10 2 5 10	DF F value 2 5.33 5 133.67 10 5.33 2 5 10	$\begin{array}{c c c} Survival \\ \hline F & P \\ \hline value & value \\ \hline 2 & 5.33 & 0.022 \\ 5 & 133.67 & <0.001 \\ 10 & 5.33 & 0.007 \\ \hline 2 \\ 5 \\ 10 \\ \hline \end{array}$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $				



ime after Salt Shock (I Fig. 6 Effect of nodulation on contents of

GSH, proline, soluble protein and soluble sugar in alfalfa leaf and root under salt shock (Data are means \pm SE of four experiments. Different letters indicate significant difference (p<0.05) amount AN, IN and NN plants at the same time point.)

Conclusion

point.)

AN

IN

Alfalfa with active nodules showed higher survival rate. Higher survival rate was associated with reduced lipid peroxidation, higher activities of superoxide dismutase (SOD), catalase (CAT), peroxidase (POD) and ascorbate peroxidase (APX) as well as higher concentrations of reduced glutathione (GSH), and soluble sugar, especially in roots under salt stress.

Methods

Seeds of alfalfa (Medicago sativa, Ladak+) were surface-sterilized and germinated in Petri dishes in a growth chamber for 5 days. Seedlings were transferred to conical plastic pots containing sterilized quartz sand without added nutrients and cultured in the greenhouse. Seedlings with 10 cm shoot height were selected and randomly divided into 3 groups: (1) inoculated with *Rhizobium meliloti* and watered daily with 1/4 strength nitrogen-free Hoagland nutrient solution, which resulted in development of active nodules (AN); (2) inoculated and watered with 1/4 strength Hoagland nutrient solution daily, which led to development of inactive nodules (IN) due to inhibition of rhizobia by sufficient nitrogen in the nutrient solution; (3) not inoculated and watered with 1/4 strength Hoagland nutrient solution, which led to plants with no nodules (**NN**). Shoots were cut on the sixtieth day and ninetieth day after inoculation to promote root and/or nodule growth. After 120 days of growth, plants were subjected to salt stress for survival tests and physiological/biochemical analyses.

and S are the abbreviation of nodulation and salt shock, respectively

Table 2 Variance analysis of contents of O_2^{-1} , MDA, GSH, proline, soluble sugar and soluble protein in the leaf and root of alfalfa under different treatments of nodulation and salt shock

Sources	O_2^{-}		MDA		G	GSH		Proline		Soluble sugar		Soluble protein	
(factors)	DF	F	Р	F	Р	F	Р	F	Р	F	Р	F	Р
(lactors)		value	value	value	value	value	value	value	value	value	value	value	value
Leaf													
N	2	44.79	< 0.001	4.11	0.022	8.64	0.001	6.02	0.004	0.82	0.446	3.11	0.053
S	5	2.93	0.021	14.16	< 0.001	12.76	< 0.001	10.85	< 0.001	6.44	< 0.001	5.71	< 0.001
N*S	10	22.97	< 0.001	20.27	< 0.001	7.94	< 0.001	2.20	0.031	5.60	< 0.001	13.03	< 0.001
Root													
N	2	21.08	< 0.001	22.14	< 0.001	1.84	0.168	0.04	0.962	26.17	< 0.001	3.75	0.030
S	5	6.29	< 0.001	5.27	0.001	8.57	< 0.001	7.70	< 0.001	3.58	0.007	3.54	0.008
N*S	10	4.99	< 0.001	6.71	< 0.001	6.78	< 0.001	5.77	< 0.001	3.94	< 0.001	3.54	0.001





Variance analysis indicated nodulation affected the activities of SOD, CAT, POD and APX along with concentrations of GSH, soluble sugar and soluble protein. Inoculation also resulted in higher basal levels of superoxide anion radical (O_2^{-1}) without salt stress.

Rhizobium symbiosis had a positive effect on alfalfa salt tolerance by improving the activity of antioxidant enzymes and osmotic adjustment capacity.

Acknowledgements

This work was supported by the Project of National Natural Science Foundation of China (31372357, 31272490), the National Key Technology R&D Program in the 12th Five-Year Plan of China (2011BAD17B05), the major project for Tibetan forage industry (Z2014C02N02) and China Agriculture Research System (CARS-35). The authors thank Drs. Yajun Wu and Roger N Gates from South Dakota State University for their advice on statistical analysis and constructive comments to this project.

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